### **Product Information**

## **Membrane Protein Extraction Kit**

Catalog Number: MeP-02

# **Description:**

This kit is designed for extracting the native, non-denatured membrane proteins from various cell types or tissues, prepared for EMSA, ELISA, 1D and 2D electrophoresis, Western blotting, TF-TF interaction arrays and other protein/DNA assays.

#### Kit contains:

Components	Quantity (50 extracts)	Storage
Cytoplasmic Lysis Buffer (C207020, blue sticker)	25.0 mL	2-8°C
Cytoplasmic Washing Buffer (C207030 purple sticker)	5.0 mL	2-8°C
Nuclear Lysis Buffer (N207040, green sticker)	2.5 mL	2-8°C
Membrane Lysis Buffer (M207060, yellow sticker)	5.0 mL	2-8°C
DTT, 1M (Dissolved in 0.1 ml ddH <sub>2</sub> O)	1 vial	-20°C
Protease/Phosphatase Inhibitors (I208052) supplied in DMSO, contains optimized AEBSF, Aprotinin, E64, Leupeptin, Pe	1 vial pstatin A, Sodium fluoride, Sodium orthovanadate a	-20°C nd Sodium pyrophosphate.

# **Protocol:** (Keep all buffers and cell/tissue samples on ice)

\*Prepare working reagents prior to proceeding.

For 10 Extractions: (50-100mg tissues or 10e7 cells/Extraction)		
Cytoplasmic Lysis Buffer (5ml)	add 5.0ul (1M DTT ) and 100ul Protease/phosphatase Inhibitors (1208052)	
Cytoplasmic Washing Buffer (1ml)	add 1.0ul (1M DTT ) and 20ul Protease/phosphatase Inhibitors (1208052)	
Nuclear Lysis Buffer (0.5ml)	add 1.0ul (1M DTT ) and 10ul Protease/phosphatase Inhibitors (1208052)	
Membrane Lysis Buffer (1ml)	add 1.0ul (1M DTT ) and 20ul Protease/phosphatase Inhibitors (1208052)	

## 1. Preparation of samples from culturing cells or tissues:

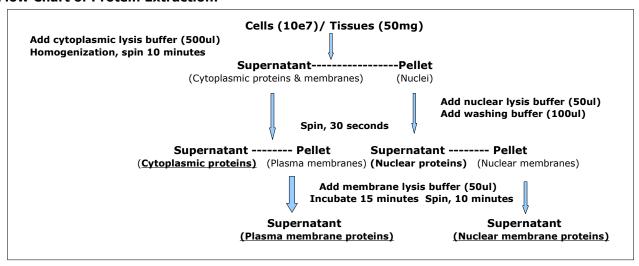
- Harvest cells (1x 10e7 cells) as usual and wash cells once with 1.0ml 1x ice-cold PBS/DPBS, centrifuge at 1,600 rpm for 8 minutes, aspirate liquids. Add 500ul cytoplasmic lysis buffer to resuspend cell pellet. Gently pipette up and down several times and incubate on ice for 10 minutes.
- Weigh 50-100mg frozen/ fresh tissues and chop tissues into small pieces using a clean razor blade. Immediately transfer into a 2.0ml microcentrifuge tube contained <u>500ul cytoplasmic lysis buffer</u>. Vortex at mid-speed for 20 seconds and incubate on ice for 10 minutes.

### 2. Homogenization:

- Using a clean pre-chilled Teflon pestle homogenizer to homogenize the cells/tissues for 10-20 strokes on ice, simply spin down the cells/tissue suspension and continue to homogenize cells/tissues another 10-20 strokes.
- (Alternative-1): Prepare a syringe with a needle gauged between 23 and 25. Pass cells/tissues through needle about 20 times to disrupt the cell membrane and release the intact nuclei and organelles.
- (Alternative-2): Using a pre-chilled, clean Dounce homogenizer to homogenize the cells/tissues twice at speed 4 (moderate) speed for 20 seconds.
- 3. Centrifuge the homogenate at 3000 rpm for 10 minutes at 4°C. Keep the pellet on ice, prepared for nuclear membrane protein extraction at **step7**.
- 4. Transfer the supernatant (containing the cytosolic, plasma membranes) to a new 1.5ml microcentrifuge. Centrifuge at 14,000xg for 30 seconds at 4°C. Aspirate liquids.
- 5. Add <u>50ul membrane lysis buffer</u> and incubate on ice for 15 minutes, Vortex vigorously 10 seconds every 5 minutes.
- 6. Centrifuge at 14,000 xg for 10 minutes at 4°C. Transfer the supernatant to a new 1.5ml microcentrifuge tube and keep on ice. This is the plasma membrane protein fractions.

- 7. Resuspend the pellet from **step 3** with <u>50ul nuclear lysis buffer</u> and vortex vigorously for 10 seconds. Incubate the suspension for 10 minutes on ice (vortex 10 seconds every 5 minutes).
- 8. Centrifuge at 14000 xg for 30 seconds at 4°C. Aspirate liquids.
- 9. Add <u>100ul cytoplasmic washing buffer</u> to resuspend the pellet. Centrifuge at 14,000 xg for 30 seconds at 4°C. Aspirate liquids. (The remained nuclear fractions were washed out).
- 10. Resuspend the pellet in <u>50ul membrane lysis buffer</u> and incubate suspension for 20 minutes on ice (vortex 10 seconds every 5 minutes).
- 11. Centrifuge at 14000 xg for 15 minutes at 4°C. Transfer the supernatant to a new 1.5ml microcentrifuge tube. This is the nuclear membrane protein fractions.
- 12. Combine nuclear membrane proteins from **step 11** and plasma membrane proteins from **step 6** to obtain total membrane protein fractions if need.
- 13. Determine the protein concentrations of plasma membrane and nuclear membrane by Bradford or BCA Assay. Stored all proteins at -80°C.

### Flow Chart of Protein Extraction:



## **Additional information:**

- The nuclear protein markers: Lamin B (68kDa), LaminA/C (70 KDa), HDAC, Histone H1 (33KDa), Histone H4(43KDa);
- The cytoplasmic protein markers: GAPDH, anti-b-actin:
- The membrane protein markers: EGFR, Na+/K+ ATPase, anti-Sp1;
- The cytoskeleton protein markers: Vimentin.
- The lysosome protein markers: LAMP1/2/3. Capthepsin D.
- The peroxisome protein markers: PMP70.
- The Zmtech protease/phosphatase Inhibitors (I208052) supplied in DMSO, contains optimized AEBSF, Aprotinin, E64, Leupeptin, Pepstatin A, Sodium fluoride, Sodium Orthovanadate and Sodium pyrophosphate.